

RESEARCH

Open Access



Assessing the potential for improved predictive capacity of antimicrobial resistance in outpatient *Staphylococcus aureus* isolates using seasonal and spatial antibiograms

Margaret Carrel^{1,4*}, Qianyi Shi^{2,3}, Gosia S. Clore^{2,3}, Shinya Hasegawa^{2,3}, Matthew Smith^{2,3}, Eli N. Perencevich^{2,3} and Michihiko Goto^{2,3}

Abstract

Background While the use of cumulative susceptibility reports, antibiograms, is recommended for improved empiric therapy and antibiotic stewardship, the predictive ability of antibiograms has not been well-studied. While enhanced antibiograms have been shown to better capture variation in susceptibility profiles by characteristics such as infection site or patient age, the potential for seasonal or spatial variation in susceptibility has not been assessed as important in predicting likelihood of susceptibility.

Methods Utilizing *Staphylococcus aureus* isolates obtained in outpatient settings from a nationwide provider of care, the Veterans Health Administration, and a local provider of care, the University of Iowa Hospitals and Clinics, standard, seasonal and spatial antibiograms were created for five commonly used antibiotic classes: cephalosporins, clindamycin, macrolides, tetracycline, trimethoprim/sulfamethoxazole.

Results A total of 338,681 *S. aureus* isolates obtained in VHA outpatient settings from 2010 to 2019 and 6,817 isolates obtained in UIHC outpatient settings from 2014 to 2019 were used to generate and test antibiograms. Logistic regression modeling determined the capacity of these antibiograms to predict isolate resistance to each antibiotic class. All models had low predictive capacity, with areas under the curve of < 0.7.

Conclusions Standard antibiograms are poor in predicting *S. aureus* susceptibility to antibiotics often chosen by clinicians, and seasonal and spatial antibiograms do not provide an improved tool in anticipating non-susceptibility. These findings suggest that further refinements to antibiograms may be necessary to improve their utility in informing choice of effective antibiotic therapy.

Keywords Antibiogram, *Staphylococcus aureus*, Seasonal, Spatial, Susceptibility

*Correspondence:

Margaret Carrel
margaret-carrel@uiowa.edu

¹Department of Geographical & Sustainability Sciences, University of Iowa, Iowa City, IA, USA

²Department of Internal Medicine, University of Iowa, Iowa City, IA, USA

³Center for Access & Delivery Research and Evaluation (CADRE), Iowa City Veterans Affairs Health Care System, Iowa City, IA, USA

⁴Iowa City, USA

© The Author(s) 2024. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

Background

As antimicrobial resistance (AMR) increases in all healthcare settings, including outpatient clinics, it is increasingly important to provide accurate information about the risk of AMR to prescribing clinicians. Cumulative susceptibility testing results, or antibiograms, is a commonly used epidemiologic method to track trends in drug resistance prevalence, guide antimicrobial empiric therapy and improve antibiotic stewardship [1–3]. Known limitations associated with antibiograms include wide variability in how antibiograms are prepared and varying confidence and ability by clinicians to interpret antibiogram information [4–12]. What is less well known, however, is whether antibiograms accurately predict resistance when empiric therapeutic decisions are made; research examining the diagnostic accuracy of antibiograms for Enterobacteriaceae isolates in a nationwide cohort in the United States (US) suggests their guidance may be poor [13].

In the US, the recommendations for antibiograms set forth by the Clinical & Laboratory Standards Institute (CLSI) include antibiogram construction at least annually and reporting results only for ≥ 30 isolates per species aggregated by facility [14, 15]. Enhanced antibiograms, where results are stratified by patient characteristics (e.g., age), culture site (e.g., urine or blood), and location (e.g., inpatient or outpatient), have been proposed to improve accuracy in guiding empiric therapy decisions, though these stratifications can result in samples sizes below the ≥ 30 isolate minimum or prove challenging for some facilities to generate [16–23].

The prevalence of bacterial infections and rates of antibiotic resistance vary seasonally and geographically in the US and globally [24–29]. Standard antibiograms do not consider the potential impacts of seasonality on AMR patterns or the varying geographic locations from which patients are drawn. Many outpatient clinics are distant from affiliated medical centers where they aggregate data, masking potential variation in environmental exposures or local circulation of resistance types, or are not affiliated with any hospital at all, meaning that AMR patterns in outpatient settings are not summarized for clinicians. Creating geography- or season-specific antibiograms could potentially improve the quality of information available to clinicians and thus improve empiric therapy decisions and antibiotic stewardship.

Utilizing curated pre-pandemic microbiology test results from the only integrated nationwide healthcare system in the US, the Veterans Health Administration (VHA), we compare the predictive accuracy of seasonal and spatial antibiograms to standard antibiograms for *Staphylococcus aureus* infections observed in outpatient settings. We also analyzed data from a regional provider of care, the University of Iowa Hospitals

& Clinics (UIHC), to test the generalizability of the approach beyond VHA data.

Methods

The VHA's Corporate Data Warehouse (CDW) was queried for all *S. aureus* clinical specimens from outpatient settings from 1/1/2010–12/31/2019. Isolates were included from all sources and sites, not only for invasive infections, given the outpatient nature of the cohort. Specimens were then excluded if they were missing geographic information (e.g., address, longitude/latitude, or county of residence) if they were obtained from a patient residing outside of the conterminous 48 states or Washington, DC (CONUS) if they were from a patient < 18 years of age or if they were obtained > 48 h after admission to institutionalized settings (e.g., nursing home, acute care) and < 72 h after discharge (see Additional Fig. 1). Clinical specimen records were then linked to microbiological results; if these susceptibility results were missing, the record was excluded. Microbiology reports were used to assess susceptibility or resistance; intermediate resistance was classified as resistant. Similar exclusion criteria were applied to *S. aureus* clinical specimen records extracted from the Epic electronic health records system of the UIHC from 1/1/2014–12/31/2019 (see Additional Fig. 2). Data from 2010 to 2014 was not available from UIHC due to changes in recordkeeping that occurred. UIHC *S. aureus* specimens from pediatric patients were retained to more accurately reflect patient distributions in US healthcare facilities than is observed in the VHA cohort and since standard antibiograms do not stratify by patient age.

For each of the datasets, three sets of antibiograms were generated for five antibiotic classes commonly used in outpatient settings: 1st–4th generation cephalosporins, clindamycin, macrolides, tetracycline and trimethoprim/sulfamethoxazole (TMP/SMX). Cephalosporin resistance is a surrogate for methicillin resistance (MRSA) as most clinical microbiology laboratories within the VHA system do not perform phenotypic susceptibility testing for methicillin or oxacillin anymore, rather they use susceptibility to 1st generation cephalosporin or ceftiofur to determine MRSA vs. MSSA. For standard antibiograms, the proportion of *S. aureus* isolates, one per patient per year, susceptible to each of the five classes was calculated in each year for each of the 138 outpatient facilities of the VHA and for the single UIHC facility. For seasonal antibiograms, susceptibility results were aggregated into four three-month seasons (December, January, February; March, April, May; June, July, August; September, October, November) across the ten years of the study, resulting in potentially forty antibiograms per facility for the VHA data and twenty-four antibiograms for the single UIHC facility. Only the first *S. aureus* isolate per patient

in each season/year was used. Spatial antibiograms were generated based upon county residence of the patient at an annual scale using the first patient record per year in each county. The potential maximum number of antibiograms is thus ten for each of the >3000 CONUS counties where *S. aureus* isolates were obtained from US veterans and five for each of the 100 counties where *S. aureus* isolates were obtained from UIHC patients.

To be consistent with the CLSI recommendations to include only routinely tested antimicrobial agents, antibiograms were generated only for antibiotic classes where susceptibility was reported for >90% of isolates per year [14]. This results in varying numbers of standard, seasonal and spatial antibiograms. To further follow CLSI recommendations, we also generated county-level, facility/year and season/year antibiograms only when 30 or more eligible isolates were available for analysis and assessed their predictive capacity separately from the entire dataset [14].

The predictive power of seasonal or spatial antibiograms versus standard antibiograms was assessed via logistic regression modeling. The modeled outcome was whether an isolate was resistant to a certain antibiotic (e.g., clindamycin), while the independent variable was the prevalence of susceptible isolates reported in the antibiogram at predefined thresholds. Thresholds of susceptibility that were examined were <80%, <85%, <90%, <95% and <98%, based upon surveys of infectious disease specialists that indicated altered decision-making in prescribing choices at susceptibility thresholds of 85–95% [30]. For example, the likelihood of clindamycin non-susceptibility in a *S. aureus* isolate collected in October 2018 from a patient residing in Johnson County, Iowa seen at UIHC would be predicted by the prevalence of clindamycin susceptibility in *S. aureus* isolates observed at UIHC in 2017, in July/August/September of 2018 and in Johnson County in 2017 at thresholds of AMR from

2 to 20%. Predictive capacity is summarized by the area under the curve (AUC); AUC>0.9 indicate high predictive capacity, 0.8–0.9 are good, 0.7–0.8 are moderate, and AUC<0.7 are poor. Sensitivity and specificity were calculated at each threshold for each model.

In addition to assessing predictive capacity, we also used VHA data to assess how many patients have county-level antibiograms available. We calculated annual proportions of patients who had county-level antibiograms available for their county of residence from the previous time frame (e.g., previous calendar year) among all patients who utilized VHA care during the respective years. This provides insight into the generalizability of this approach in counties with smaller populations.

All statistical analyses were performed with R version 4.3.1 (R Foundation for Statistical Computing, Vienna, Austria). Ethical approval for the study was obtained from the Iowa City Veterans Affairs Health Care System and University of Iowa Institutional Review Boards.

Results

A total of 338,681 *S. aureus* isolates obtained in VHA outpatient settings from 2010 to 2019 and 6,817 isolates obtained in UIHC outpatient settings from 2014 to 2019 were used to generate and test antibiograms. Patient demographics from VHA differed from that of UIHC: the VHA dataset was majority male while the UIHC data was more evenly split according to gender, and the UIHC dataset had approximately one-fifth of samples from pediatric patients while these were excluded from the VHA dataset ($n=2$) (Table 1).

Though the number varied by antibiotic class and temporally, VHA isolates were drawn from a range of 84–136 of the 138 outpatient VHA facilities and from 2337 to 2836 of the 3,038 counties in the CONUS with outpatient veteran visits (Table 2). UIHC data came from a single facility and 98–100 counties, predominantly in Iowa and Western Illinois. When the data was subset to consider only facilities or counties with ≥ 30 isolates, the majority of counties and isolates in the VHA data were excluded, though this varied by antibiotic class. All UIHC *S. aureus* samples were tested against all five antibiotic classes but only ten counties met the ≥ 30 isolate minimum.

Antibiotic resistance rates for some antibiotic classes differed between the VHA and UIHC datasets. Cephalosporin resistance was over 10% higher in VHA *S. aureus* samples than in UIHC, and macrolides resistance was 5% higher. Clindamycin resistance in UIHC samples was 8% higher than in VHA *S. aureus*. Resistance rates for tetracyclines and TMP/SMX were comparable.

All antibiograms performed poorly in predicting resistance, across all five antibiotic classes, with AUC below 0.7 (Table 3). Including only facilities, counties or seasons with ≥ 30 isolates did not significantly improve predictive

Table 1 Demographic characteristics of patients with positive *S. aureus* outpatient cultures in the VHA and UIHC.

Variable	VHA		UIHC	
	N	%	N	%
Total	338,681		6817	
Gender				
Female	18,084	5.3%	3151	46.2%
Male	320,597	94.7%	3666	53.8%
Age				
< 18	0	0.0%	1564	22.9%
18–35	16,950	5.0%	1295	19.0%
35–50	33,701	10.0%	1054	15.5%
50–65	122,536	36.2%	1540	22.6%
65–80	120,678	35.6%	1035	15.2%
80+	44,816	13.2%	329	4.8%

VHA: Veterans Health Administration; UIHC: University of Iowa Hospitals & Clinics

Table 2 Resistance rates of *S. aureus* to antimicrobial classes in the VHA and UIHC and samples sizes for facilities and counties included in antibiogram generation for the overall datasets and then the subset that met the ≥ 30 isolate CLSI recommendation

Antibiotic Group	VHA				UIHC					
	Overall Resistance	Facilities	Facilities ≥ 30	Counties	Counties ≥ 30	Overall Resistance	Facilities	Facilities ≥ 30	Counties	Counties ≥ 30
Cephalosporins	44.31%	136	130	2836	328	33.15%	1	1	100	10
Clindamycin	23.82%	85	79	2505	201	32.51%	1	1	98	10
Macrolides	59.11%	84	78	2337	194	54.60%	1	1	98	10
Tetracyclines	6.11%	128	123	2686	299	7.45%	1	1	100	10
TMP/SMX	3.09%	130	125	2627	309	3.54%	1	1	100	10

VHA: Veterans Health Administration; UIHC: University of Iowa Hospitals & Clinics

Table 3 Performance metrics for standard, seasonal and spatial antibiograms generated for five antimicrobial classes for overall datasets and then those facilities/counties that met the ≥ 30 isolate CLSI recommendation

	Antibiotic	Standard	Seasonal	Spatial
VHA	Cephalosporins	0.581	0.577	0.563
	Clindamycin	0.554	0.540	0.543
	Macrolides	0.543	0.541	0.541
	Tetracyclines	0.571	0.553	0.551
	TMP/SMX	0.653	0.641	0.630
VHA ≥ 30	Cephalosporins	0.583	0.573	0.571
	Clindamycin	0.550	0.543	0.539
	Macrolides	0.551	0.543	0.543
	Tetracyclines	0.564	0.544	0.555
	TMP/SMX	0.657	0.641	0.646
UIHC	Cephalosporins	0.513	0.495	0.552
	Clindamycin	0.504	0.507	0.529
	Macrolides	0.514	0.530	0.539
	Tetracyclines	0.550	0.515	0.564
	TMP/SMX	0.578	0.533	0.616
UIHC ≥ 30	Cephalosporins	0.513	0.496	0.534
	Clindamycin	0.504	0.507	0.558
	Macrolides	0.513	0.530	0.571
	Tetracyclines	0.550	0.515	0.559
	TMP/SMX	0.578	0.533	0.651

VHA: Veterans Health Administration; UIHC: University of Iowa Hospitals & Clinics

ability. Standard antibiograms had the greatest predictive capacity for VHA datasets while spatial antibiograms had the greatest predictive capacity for UIHC datasets. Seasonal, facility level antibiograms performed the worst in all trials. Higher, though still very poor, predictive ability was observed for antibiotic classes with the highest levels of susceptibility. This is reflected in reasonable sensitivity and specificity levels at high (>90%) thresholds only for tetracyclines and TMP/SMX and only in the VHA models with large enough sample sizes (Additional Table 1).

The proportion of patients who would have annual county-level antibiograms from the previous calendar year to use in predicting susceptibility among the entire patient population for each calendar year ranged from 44.7 to 49.0% (mean: 46.8%).

Discussion

In the analysis of the potential for seasonal and spatial antibiograms to improve the capacity to predict non-susceptibility of *S. aureus* to five antibiotic classes, models built using >330,000 isolates from patients across the US seen in outpatient VHA facilities and using >6000 isolates from patients seen in outpatient UIHC facilities performed poorly, on par with flipping a coin. Antibiograms created according to current CLSI standards for each facility (ignoring geographic information) were similarly inaccurate in predicting *S. aureus* susceptibility. The poor performance of standard antibiograms using VHA data has previously been observed for Enterobacteriaceae [13]. While UIHC’s patient population is less spatially representative than VHA, it is more balanced in terms of gender and age, but antibiograms generated using this more diverse patient dataset still performed poorly.

When the ≥ 30 minimum was implemented, the number of counties, and by extension isolates, included in the analysis declined greatly for both the VHA and UIHC dataset and the coverage of the total veteran population residing in included counties fell from 95 to 47%. Thus, even for the largest provider of healthcare in the US, generating spatial antibiograms while following the CLSI recommendation only to use data with 30 or more isolates is difficult. The creation of community-level antibiograms, ranging from facilities in a single county to multi-county, has been suggested as a tool for facilities that may otherwise fail to meet the ≥ 30 isolate minimum [31, 32]. This, however, poses a challenge for greater granularity in considering the social and environmental characteristics of places where patients reside and how this can impact the susceptibility profiles of their infections. Determining best practices to meet the CLSI recommended isolate numbers is necessary; for example, using spatial smoothing or aggregating facilities/counties based upon shared sociodemographic or environmental characteristics rather than adjacency might improve the predictive capabilities of antibiograms.

The strength of this study includes the use of a large cohort from the only integrated healthcare system with a presence in all conterminous states in the US, the VHA,

with more than 300,00 isolates. The standardized data extraction process and aggregation methods, rather than reliance on locally produced antibiograms, can avoid the problem from the practice variation in antibiogram preparation. A previous study indicated that only a small fraction of facilities are completely adhering to the CLSI guidelines [10]. Studies that assessed antibiogram generation and usage in other nations or regions are sparse, but some suggested this situation might be similar in European nations where guidelines published by the European Committee on Antimicrobial Susceptibility Testing (EUCAST) are used [33].

There are several limitations to this study. First, the VHA provides care to more elderly, male-dominant populations compared to the nationwide demographics of the US. Previous studies suggested that males have a higher risk of methicillin-resistant *S. aureus* (MRSA) carriage and invasive infections, but data for gender differences in resistance prevalence for other antimicrobial classes are sparse [34]. This may potentially limit the generalizability to populations outside the VHA. However, while the proportion of females has increased gradually over the years, there has been no drastic and sudden change during the study period [35]. Additionally, the UIHC cohort more closely resembles the age and gender profile of the US and models using this dataset performed similarly to the VHA data. Second, we focused on assessing standardized antibiograms based on the CLSI guidelines and did not compare them with antibiograms generated by other standards, such as the EUCAST guidelines.

Conclusions

Although seasonal and spatial antibiograms performed as or more poorly than standard antibiograms, spatio-temporal variation in infection prevalence and antibiotic resistance coupled with observed variation in antibiograms that incorporate patient-level and other information suggest that there is utility in learning how to build better antibiograms [18, 20, 22, 25–27, 29]. Future work will merge information on patients with known characteristics of their infections and data from the communities they were drawn to develop models to better predict antibiotic susceptibility. This, coupled with ongoing work in revising how antibiograms are visualized and improving their useability, means that antibiograms may yet become the tools to guide empiric therapy and improve antibiotic stewardship for which they are currently recommended [8, 36–38].

Abbreviations

AMR	Antimicrobial resistance.
CDW	Corporate Data Warehouse.
CLSI	Clinical & Laboratory Standards Institute.
CONUS	Conterminous US (48 states and Washington, DC).
EUCAST	European Committee on Antimicrobial Susceptibility Testing.
MRSA	Methicillin-resistant <i>S. aureus</i> .

TMP/SMX	Trimethoprim/Sulfamethoxazole.
UIHC	University of Iowa Hospitals & Clinics.
US	United States.
VHA	Veterans Health Administration.

Supplementary information

The online version contains supplementary material available at <https://doi.org/10.1186/s13756-024-01388-3>.

Supplementary additional table. 1. Sensitivity (sn) and specificity (sp) of standard, seasonal and spatial antibiograms predicting non-susceptibility of *S. aureus* isolates at predefined thresholds.

Supplementary additional fig. 1. The creation process of standardized, spatial and seasonal antibiograms for the VHA dataset.

Supplementary additional fig. 2. The creation process of standardized, spatial and seasonal antibiograms for the UIHC dataset.

Acknowledgements

We would like to thank Jonas Church for his assistance.

Author contributions

MC & MG conceived of the study design; MC, MS, GC, SH, QS & MG conducted data acquisition, preparation and analysis; MC, QS & MG drafted the work; SH, MS, QS, GC & ENP revised the work; all authors gave approval of the final version.

Funding

MC, QS & GC received support from the Centers for Disease Control & Prevention Epicenter program award CK000613. MG received support from the Agency for Healthcare Research and Quality under award number K08HS027472. ENP received support from the Veterans Affairs Health Services Research and Development (HSR&D) Service Center of Innovation under award number 13–412. MS and SH report no relevant funding sources. The views expressed in this article are those of the authors and do not necessarily reflect the position or policy of the Department of Veterans Affairs or the United States Government. The funders have no role in the design and conduct of the study; collection, management, analysis, and interpretation of the data; preparation, review, or approval of the manuscript; or decision to submit the manuscript for publication.

Data availability

The datasets generated in this study are not publicly available due to their sensitive nature and ownership by the VHA and UIHC. Access to the datasets may be granted by VHA or UIHC.

Declarations

Ethics approval and consent to participate

The study was approved by the Veterans Administration and University of Iowa Institutional Review Boards (#202109177 & #202308251).

Consent for publication

Not applicable.

Competing interests

The authors declare no competing interests.

Received: 22 January 2024 / Accepted: 11 March 2024

Published online: 22 March 2024

References

1. Fridkin SK, Edwards JR, Tenover FC, Gaynes RP, McGowan JE Jr. Intensive care Antimicrobial Resistance Epidemiology P, National Nosocomial Infections Surveillance System H. Antimicrobial resistance prevalence rates in hospital antibiograms reflect prevalence rates among pathogens associated with

- hospital-acquired infections. *Clin Infect Diseases: Official Publication Infect Dis Soc Am.* 2001;33(3):324–30.
2. Goff DA. Antimicrobial stewardship: bridging the gap between quality care and cost. *Curr Opin Infect Dis.* 2011;24(Suppl 1):S11–20.
 3. Toth NR, Chambers RM, Davis SL. Implementation of a care bundle for antimicrobial stewardship. *Am J Health-System Pharm.* 2010;67(9).
 4. Salsgiver E, Bernstein D, Simon MS, Eiras DP, Greendyke W, Kubin CJ, et al. Knowledge, attitudes, and practices regarding antimicrobial use and stewardship among prescribers at acute-care hospitals. *Infect Control Hosp Epidemiol.* 2018;39(3):316–22.
 5. Cooper SJ, Destache CJ, Vivekanandan R. Improving understanding and utilization of the antibiogram among medical residents. *Antimicrob Stewardship Healthc Epidemiol.* 2022;2(1):e142.
 6. Mermel LA, Jefferson J, Devolve J. Knowledge and use of cumulative antimicrobial susceptibility data at a university teaching hospital. *Clin Infect Dis.* 2008;46(11):1789.
 7. Tallman GB, Vilches-Tran RA, Elman MR, Bearden DT, Taylor JE, Gorman PN, McGregor JC. Empiric antibiotic prescribing decisions among medical residents: the role of the antibiogram. *Infect Control Hosp Epidemiol.* 2018;39(5):578–83.
 8. Truong WR, Hidayat L, Bolaris MA, Nguyen L, Yamaki J. The antibiogram: key considerations for its development and utilization. *JAC-antimicrob Resist.* 2021;3(2):dlab060.
 9. Simner PJ, Hindler JA, Bhowmick T, Das S, Johnson JK, Lubers BV, et al. What's New in Antibiograms? Updating CLSI M39 Guidance with current trends. *J Clin Microbiol.* 2022;60(10):e02210–21.
 10. Moehring RW, Hazen KC, Hawkins MR, Drew RH, Sexton DJ, Anderson DJ. Challenges in preparation of cumulative antibiogram reports for community hospitals. *J Clin Microbiol.* 2015;53(9):2977–82.
 11. Xu R, Polk RE, Stencel L, Lowe DK, Guharoy R, Duggal RW, et al. Antibiogram compliance in University HealthSystem Consortium participating hospitals with Clinical and Laboratory Standards Institute guidelines. *Am J Health-System Pharm.* 2012;69(7):598–606.
 12. Lautenbach E, Nachamkin I. Analysis and presentation of cumulative antimicrobial susceptibility data (antibiograms): substantial variability across medical centers in the United States. *Infect Control Hosp Epidemiol.* 2006;27(4):409–12.
 13. Hasegawa S, Livorsi DJ, Perencevich EN, Church JN, Goto M. Diagnostic Accuracy of Hospital Antibiograms in Predicting the risk of Antimicrobial Resistance in Enterobacteriaceae isolates: a nationwide Multicenter evaluation at the Veterans Health Administration. *Clin Infect Dis.* 2023;77(11):1492–500.
 14. Clinical, Laboratory Standards I. Analysis and Presentation of Cumulative Antimicrobial Susceptibility Test Data; Approved Guideline-Third Edition (M39-A3). Wayne, PA: Clinical and Laboratory Standards Institute (CLSI); 2009. Contract No.: Report.
 15. Hindler JF, Stelling J. Analysis and presentation of cumulative antibiograms: a new consensus guideline from the Clinical and Laboratory standards Institute. *Clin Infect Diseases: Official Publication Infect Dis Soc Am.* 2007;44(6):867–73.
 16. Tran C, Hargy J, Hess B, Pettengill MA. Estimated impact of low isolate numbers on the Reliability of Cumulative Antibiogram Data. *Microbiol Spectr.* 2023;11(1):e03939–22.
 17. Pogue JM, Alaniz C, Carver PL, Pleva M, Newton D, DePestel DD. Role of unit-specific combination antibiograms for improving the selection of appropriate empiric therapy for gram-negative pneumonia. *Infect Control Hosp Epidemiol.* 2011;32(3):289–92.
 18. Grodin L, Conigliaro A, Lee S-Y, Rose M, Sinert R. Comparison of UTI antibiograms stratified by ED patient disposition. *Am J Emerg Med.* 2017;35(9):1269–75.
 19. Jorgensen S, Zurayk M, Yeung S, Terry J, Dunn M, Nieberg P, Wong-Beringer A. Emergency department urinary antibiograms differ by specific patient group. *J Clin Microbiol.* 2017;55(9):2629–36.
 20. McGregor JC, Bearden DT, Townes JM, Sharp SE, Gorman PN, Elman MR, et al. Comparison of antibiograms developed for inpatients and primary care outpatients. *Diagn Microbiol Infect Dis.* 2013;76(1):73–9.
 21. Rabs N, Wiczorkiewicz SM, Costello M, Zamfirova I. Development of a urinary-specific antibiogram for gram-negative isolates: impact of patient risk factors on susceptibility. *Am J Infect Control.* 2014;42(4):393–400.
 22. Swami SK, Banerjee R. Comparison of hospital-wide and age and location-stratified antibiograms of *S. Aureus*, *E. Coli*, and *S. pneumoniae*: age- and location-stratified antibiograms. *Springerplus.* 2013;2(1):1–5.
 23. Tiwari K, Patil S, Naik A, Shetty A, Walia K, Rodrigues C. Patient risk factor stratification is essential for the hospital antibiogram. *Int J Infect Control.* 2021;17.
 24. Eber MR, Shardell M, Schweizer ML, Laxminarayan R, Perencevich EN. Seasonal and temperature-associated increases in gram-negative bacterial bloodstream infections among hospitalized patients. *PLoS ONE.* 2011;6(9):e25298.
 25. Leekha S, Diekema D, Perencevich E. Seasonality of staphylococcal infections. *Clin Microbiol Infect.* 2012;18(10):927–33.
 26. Livorsi DJ, Goto M, Carrel M, Jones MM, McDanel J, Nair R, et al. Regional variations in fluoroquinolone non-susceptibility among *Escherichia coli* bloodstream infections within the Veterans Healthcare Administration. *Antimicrob Resist Infect Control.* 2016;5(1):38.
 27. Perencevich EN, McGregor JC, Shardell M, Furuno JP, Harris AD, Morris JG, et al. Summer peaks in the incidences of gram-negative bacterial infection among hospitalized patients. *Infect Control Hosp Epidemiol.* 2008;29(12):1124–31.
 28. McCormick AW, Whitney CG, Farley MM, Lynfield R, Harrison LH, Bennett NM, et al. Geographic diversity and temporal trends of antimicrobial resistance in *Streptococcus pneumoniae* in the United States. *Nat Med.* 2003;9(4):424–30.
 29. Smith MW, Carrel M, Goto M. County-level spatiotemporal distribution of fluoroquinolone-resistant Enterobacteriaceae in outpatient settings of the veterans' Health Administration, 2000–2017. *Infect Control Hosp Epidemiol.* 2022:1–3.
 30. Hasegawa S, Perencevich EN, Dukes K, Goto M, editors. 964. Physicians' acceptable treatment failure rates and interpretation of antibiogram for Gram-negative infections: A pilot survey study of infectious diseases specialists. IDWeek 2023. Boston MA. Open forum infectious diseases. Oxford University Press US; 2022.
 31. Hostler CJ, Moehring RW, Ashley ESD, Johnson M, Davis A, Lewis SS, et al. Feasibility and value of developing a regional antibiogram for community hospitals. *Infect Control Hosp Epidemiol.* 2018;39(6):718–22.
 32. Woldai S, Ward M, Hughes SM, Sexton T, Tran J, Chung W, editors. Utility and limitations of an Aggregate Community Antibiogram, Dallas County, Texas, 2009–2015. Oxford University Press; 2016. Open Forum Infectious Diseases.
 33. Patel S, Jhass A, Slee A, Hopkins S, Shallcross L. Variation in approaches to antimicrobial use surveillance in high-income secondary care settings: a systematic review. *J Antimicrob Chemother.* 2021;76(8):1969–77.
 34. Humphreys H, Fitzpatrick F, Harvey BJ. Gender differences in rates of carriage and bloodstream infection caused by methicillin-resistant *Staphylococcus aureus*: are they real, do they matter and why? *Clin Infect Dis.* 2015;61(11):1708–14.
 35. Affairs DoV. The past, present and future of women veterans. Report by the national center for veterans analysis and statistics. 2017.
 36. Dauphin A, McCoy C, Bowden R, Lee M, Gold H, Chapin R. Creating an electronic antibiogram using visualization software: easily updatable and removes the need for yearly manual review. *Antimicrob Stewardship Healthc Epidemiol.* 2023;3(S2):s34–s.
 37. Simpaio AF, Ahumada LM, Martinez BL, Cardenas AM, Metjian TA, Sullivan KV, et al. Design and implementation of a visual analytics electronic antibiogram within an electronic health record system at a tertiary pediatric hospital. *Appl Clin Inf.* 2018;9(01):037–45.
 38. Vingino AR, Rabinowitz P, Kapoor H, Ramirez V, Salm A, editors. 177. User Preferences for Visualization of Antibiogram Data in Clinical Practice for Empiric Prescription of Antibiotics. Open Forum Infectious Diseases; 2021: Oxford University Press US.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.